

## Histopathology of rainbow trout gills after exposure to copper

Mahboobeh KHABBAZI<sup>1</sup>, Mohammad HARSIJ<sup>1</sup>, Seyed Ali Akbar HEDAYATI<sup>2</sup>, Mohammad Hasan GERAMI<sup>1,3\*</sup>, Hamed GHAFARI-FARSANI<sup>4</sup>

<sup>1</sup>Department of Fisheries, Gonbad Kavous University, Gonbad Kavous, Iran.

<sup>2</sup>Department of Fisheries, Gorgan University of Agricultural and Natural Resources, Gorgan, Iran.

<sup>3</sup>Young Researchers & Elite Club, Shiraz Branch of Islamic Azad University, Shiraz, Iran.

<sup>4</sup>Young Researchers & Elite Club, Shahrekord Branch of Islamic Azad University, Shahrekord, Iran.

<sup>4</sup>Young Researchers & Elite Club, Shiraz Branch of Islamic Azad University, Shiraz, Iran.

\*Email: m.h.gerami@gonbad.ac.ir

**Abstract:** Copper is a very common element in water resources. For this reason, concerns about the risks and consequences of this element in water contamination are arising. The purpose of this study was to investigate the effects of water-born CuO (Copper (II) oxide) on the rainbow trout, *Oncorhynchus mykiss* gill tissue to establish a suitable biomarker for copper in water resources. Samples with 18±3g body weight were exposed to CuO for a week with 0.0125, 0.037, 0.075, and 0.15 ppm of copper sulfate and a control group (without CuO). Physicochemical properties of water were 15±2°C, pH 7-8, CaCO<sub>3</sub> 270mg/l and oxygen saturation 90.9±0.2%. At the end of 7 days, 9 fish were caught randomly from each treatment and second gill from left side of fish were removed for histological study. Hyperplasia, oedema, epithelial lifting in secondary lamella and lamellar aneurysm were observed in gill tissues, showing that CuO, as a copper ion, has significant adverse effects on gill tissues of rainbow trout and gills can be a suitable biomarker for copper in water resources.

**Keywords:** Biomarker, Gill, Heavy metals, *Oncorhynchus mykiss*.

### Introduction

Copper (Cu) is a very common element with a ubiquitous distribution in the environment and biota (Carbonell & Tarazona 1994). Copper is an essential micronutrient naturally occurring in unpolluted fresh waters, in concentrations ranging from 0.2 to 30mg/l (USEPA 2007). Although copper is essential to animals and higher plants in low amounts (e.g., cytochrome c oxidase and superoxide dismutase) (Zhou & Gitschier 1997), hazard and pathological effects of copper is reasonably well-known in fish in excess amounts (Grosell et al. 2007; Mustafa et al. 2012). Copper causes destructive effect on the ultrastructure of the pavement cells (PVC) and chloride cells (CC) of fish gill epithelia such as lifting, rupture peeling of lamellar epithelium, lamellar fusion, hyperplasia, and cellular

hypertrophy (Pelgron et al. 1995; Dang et al. 2000; Mazon et al. 2002). However, the severity of these damages depends on the species' sensitivity, the copper concentration, and the exposure time.

Along with the development of industries, a progressive decline in the water quality occurred resulting in poor ecological condition of the water bodies. Chemical analyses may not be sufficient to properly assess the adverse effects of the complex mixture of water contaminants (He et al. 2011). Thus, to assess the destructive effects of pollutant in water bodies, a biomarker-based bio-monitoring is a promising approach to provide early warning signs of exposure (Viarengo et al. 2000; Au 2004; Zorita et al. 2007; Tlili et al. 2010; Pereira et al. 2013). According to Water Framework Directive (WFD), fish represent one of the key elements to evaluate the rivers

ecological conditions (Scardi et al. 2008; Hermoso et al. 2010). Fish gills are the first target of pollutants because of their large surface area, quick absorption and direct contact with the external environment (Pandey et al. 2008). For this reason, fish gills are suitable organs for toxicology studies.

Analyzing the histopathological changes in gill tissue has been an instrument widely used in aquatic toxicology to bio-monitor acute and chronic situations (Stentiford et al. 2003; Van Dyke & Pieterse 2008; Pereira et al. 2013). Changes in gill epithelium are a consequence of a range of contaminant exposures, with the severity of changes depending on the pollutant concentration and exposure period (Santos et al. 2011). Therefore, the aim of this study was to evaluate gill histopathology of rainbow trout after exposure to copper sulfate. Rainbow trout, *Oncorhynchus mykiss*, is an economically important species commercially farmed in many countries throughout the world. It is exotic to Iranian inland waters which introduced to the Tigris River, Caspian Sea, Lake Urmia, Namak Lake, Kavir, Esfahan and Kor River basins (Esmaili et al. 2014) and has a rapid growth that can easily adapt to environmental conditions. Finding data on environmental biomarker for this species can be useful in aquatic toxicity management and environmental safety.

## Materials and methods

One hundred and fifty-five live specimens of rainbow trout were obtained from local farms in October 2013. Samples weighted  $18 \pm 3$ g in average. They were acclimatized in an aquarium (30×20×35cm; 2100 liter) for one week in the Gonbad Kavous University Fisheries Laboratory. Fish were divided into five groups (three replicates) with 10 individuals in each aquarium and were treated with 0.00 (control group), 0.0125, 0.037, 0.075, and 0.15ppm of copper sulfate. These sub-lethal concentrations were determined according to ATSDR (1990). It is noted that ATSDR, determined lethal concentration of copper 1-8ppm for freshwaters fishes. No feed was

given during the experiment (one week). There were no significant differences between the aquaria in water quality in the following parameters: pH:  $7 \pm 0.004$ ; temperature:  $15 \pm 2^\circ\text{C}$ ; hardness:  $270 \pm 0.05$ ppm and oxygen saturation:  $90.9 \pm 0.2\%$ . The photoperiod was 12h light and 12h dark.

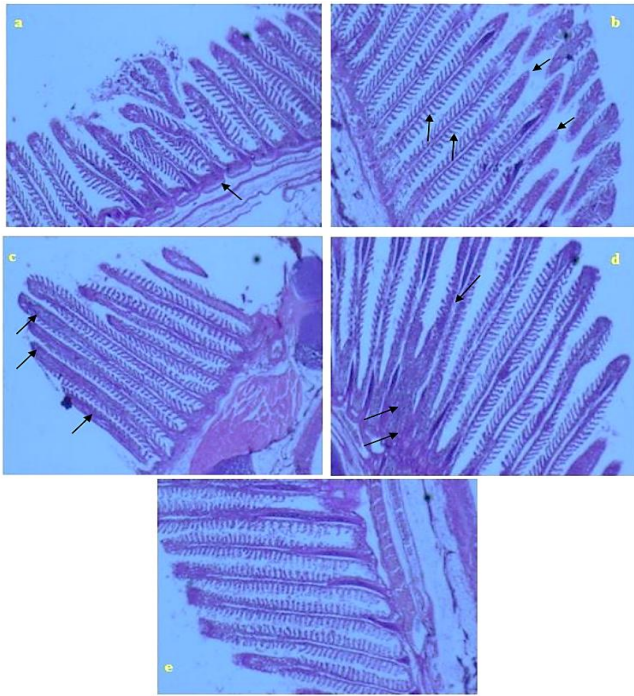
After one week, nine fish were caught randomly from each aquarium and the second gill arch from left side of each fish was separated for histopathological studies and fixed in 10% formalin solution and dehydrated in 96% ethanol, cleared with xylene, impregnated with paraffin, embedded, sectioned, mounted, and stained with Haematoxylin and Eosin (H&E) (Bucke 1972). Histopathological changes induced by treatments in the tissues were photographed using Nikon photomicroscope.

## Results

Gill morphology of rainbow trout was normal in all the unexposed control samples. Exposure to copper sulphate caused gill injuries such as Hyperplasia (shortening, rounding and fusion of the secondary lamellae), epithelial hypertrophy (lifting cell layer by oedematous), lamellar fusion (coupling between two adjacent secondary lamellar epithelium) and lamellar aneurism (shutting capillary path within the secondary lamellae) (Fig. 1). Hyperplasia and swollen mucocytes was the most observed injuries. Table 1 shows type of injuries with damage degree. The results showed that hyperplasia increased until 0.037ppm copper concentration and decreased in

**Table 1.** Rainbow trout gill damage during the lethal concentrations of copper sulfate.

Types of injuries	Sub-lethal concentrations (ppm)				
	Control	0.0125	0.037	0.075	0.15
Hyperplasia	-	+	+++	+	-
Lamellar fusion	-	-	-	+++	+
Lamellar aneurism	-	-	-	+	++
Leukocyte infiltration	-	-	+	-	+++
Epithelial hypertrophy	-	+++	++	+	+



**Fig.1.** Microphotographs of gill histopathological changes by copper sulfate in Rainbow trout (a-d) and control group (e). Epithelial hypertrophy (a), Hyperplasia (b), lamellar fusion and swollen mucocytes (c) and lamellar aneurism and oedema (d).

0.075ppm. Lamellar aneurism and leukocyte infiltration increased in 0.15ppm copper concentration while hyperplasia, lamellar fusion and epithelial hypertrophy decreased (Fig. 1).

## Discussion

Copper is a well-known respiratory and ionoregulatory toxicant in fish (Handy 2003; Grosell et al. 2007; Al-Bairuty et al. 2013). Mucus secretion is often the first line of defense to metal exposure in the gills. This phenomenon can temporarily protect gill tissue from injuries (Handy & Maunder 2009). Mucus secretion and swollen mucocytes were observed in this study, but it seems to be not sufficient to protect the gills from the injury causing by copper sulfate. Although hyperplasia and lamellar fusion known to be induced by many gill tissue irritants, however, focal points of cellular hypertrophy and necrosis followed by epithelial rupture, reflect the direct deleterious effects of heavy metals in fish gills (Mazon et al. 2002). In addition,

similar gill injuries by Titanium (TiO<sub>2</sub>) in *Cyprinus carpio* (Hao et al. 2009) and Silver in *Danio rerio* (Griffitt et al. 2009) were reported. Further, the types of gill injuries by copper evaluated in this study also have been reported by other researchers (Monteiro et al. 2008; Al-Bairuty et al. 2013; Pereira et al. 2013).

Rainbow trout gill had various reactions in different copper doses. In 0.0125ppm, slight hyperplasia and sever epithelial hypertrophy was observed. This reaction shows that the fish immune system is ready to respond to contaminant. Epithelial thickening was attributed to the appearance of macrophages and other leucocytes integrated in a compensatory response of tissue repair (Teh et al. 1997). Monteiro et al. (2008) reported that hyperplasia and epithelial hypertrophy were clearly related to low copper doses and lifting was not significantly correlated with gill copper deposition at high copper doses. They also declared that lamellar fusion occurs in chronic copper exposure which coincided with our findings.

Monteiro et al. (2008) stated that oedema and aneurisms were clearly related to short-term copper exposure, lamellar fusion with chronic copper exposure, lifting with increasing time at the lower metal concentrations and changes in filament epithelium thickness with the higher metal concentrations. According to Mallatt (1985), gill lesions can be divided into two groups; one that reflects the direct effect of toxicants and another one that corresponding to defense responses of fishes. Pereira et al. (2013) stated that filament epithelium proliferation and necrosis were the biological responses that contribute mostly to discriminate the ecological status classification. Several studies have been demonstrated, on laboratory experiments, to determine the toxicity of heavy metals, organochlorine pesticides and petroleum hydrocarbon products to fish gill (Al-Attar, 2007; Garcia-Santos et al. 2007; Patnaik et al. 2011; Santos et al. 2011; Hesni et al. 2011; Moitra et al. 2012), and many researchers used fish gill as a tool to assess the presence of contaminants, in natural aquatic systems

(Stentiford et al. 2003; Fernandes et al. 2007). Fernandes et al. (2007) previously stated that lesions severity and extension might be an accurate indicator of the toxicant levels, but the extent and severity of each particular lesion is variable among different species.

This study showed that gill biological responses could be suggested as a useful biomarker to identify ecosystems perturbation. However, reaction to water contaminations is varying for each species; but the main injuries in each group of pollutants (such as metals, pesticides, herbicides etc.) are constant. In general, each biological response reflects the degree of water contamination.

### Acknowledgments

This study was financially supported by Gonbad Kavous University and Gorgan University of Agricultural Sciences & Natural Resources. We are pleased to thank Dr. Rahman Patimar and A. Bahalkeh for their sincere assistance in conducting this research.

### References

- Al-Attar, A.M. 2007. The influences of nickel exposure on selected physiological parameters and gill structure in the teleost fish *Oreochromis niloticus*. *Journal of Biological Sciences* 7: 77-85.
- Al-Bairuty, G.A.; Shaw, B.D.; Handy, R.B. & Henry, T. 2013. Histopathological effects of waterborne copper nanoparticles and copper sulphate on the organs of rainbow trout (*Oncorhynchus mykiss*). *Aquatic Toxicology* 126: 104-115.
- ATSDR (Agency for Toxic Substances and Disease Registry). 1990. Toxicological Profile for Copper. US Public Health Service, Atlanta, Georgia. TP-90-08. 143 pp.
- Au, D.W.T. 2004. The application of histocytopathological biomarkers in marine pollution monitoring: a review. *Marine Pollution Bulletin* 48: 817-834.
- Bucke, D. 1982. Some histological techniques applicable to fish tissues, Mawdesley -Thomas. LE. Edn 10, Diseases of fish. Symposium of Zoology Society, London, Vol. 30, Academic Press, New York, 153p.
- Carbonell, G. & Tarazona, J.V. 1994. Toxicokinetics of copper in rainbow trout. *Aquatic Toxicology* 29: 213-221.
- Dang, Z.; Lock, R.A.C.; Flik, G. & Wendelaar Bonga, S.E. 1999. The metallothionein response in gills of *Oreochromis mossambicus* exposed to copper in fresh water. *American Journal of Physiology* 277: R320-R331.
- Esmaili, H.R.; Coad, B.W.; Mehraban, H.R.; Masoudi, M.; Khaefi, R.; Abbasi, K.; Mostafavi, H. & Vatandoust, S. 2014. An updated checklist of fishes of the Caspian Sea basin of Iran with a note on their zoogeography. *Iranian Journal of Ichthyology* 1(3): 152-184.
- Fernandes, C.; Fontainhas-Fernandes, A.; Monteiro, S.M. & Salgado, M.A. 2007. Histopathological gill changes in wild leaping gray mullet (*Liza saliens*) from the Esmoriz-Paramos coastal lagoon, Portugal. *Environmental Toxicology* 22: 443-448.
- Garcia-Santos, S.; Monteiro, M.; Carrola, J. & Fontainhas-Fernandes, A. 2007. Histopathological lesions of tilapia *Oreochromis niloticus* gills caused by cadmium. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia* 59: 376-381.
- Griffitt, R.J.; Hyndman, K.; Denslow, N.D. & Barber, D.S. 2009. Comparison of molecular and histological changes in zebrafish gills exposed to metallic nanoparticles. *Toxicology Sciences* 107: 404-415.
- Grosell, M.; Blanchard, J.; Brix, K.V. & Gerdes, R. 2007. Physiology is pivotal for interactions between salinity and acute copper toxicity to fish and invertebrates. *Aquatic Toxicology* 84: 162-172.
- Handy, R.D. & Maunder, R.J. 2009. The biological roles of mucus: importance for osmoregulation and osmoregulatory disorders of fish health. In: Handy, R.D., Bury, N.R. & Flik, G. (Eds.), *Osmoregulation and Ion Transport: Integrating Physiological, Molecular and Environmental Aspects. Essential Reviews in Experimental Biology*, vol. 1. Society for Experimental Biology Press, London, pp. 203-235.
- Handy, R.D. 2003. Chronic effects of copper exposure versus endocrine toxicity: two sides of the same toxicological process? *Comparative Biochemistry*

- and Physiology - Part A: Molecular & Integrative Physiology 135: 25-38.
- Hao, L.; Wang, Z. & Xing B. 2009. Effect of sub-acute exposure to TiO<sub>2</sub> nanoparticles on oxidative stress and histopathological changes in juvenile carp (*Cyprinus carpio*). Journal of Environmental Sciences 21: 1459–1466.
- He, X.; Nie, X.; Wang, Z.; Cheng, Z.; Li, K.; Li, G.; Wong, M.H.; Liang, X. & Tsui, M.T.K. 2011. Assessment of typical pollutants in waterborne by combining active biomonitoring and integrated biomarkers response. Chemosphere 84: 1422-1431.
- Hermoso, V.; Clavero, M.; Blanco-Garrido, F. & Prenda, J., 2010. Assessing the ecological status in species-poor systems: a fish-based index for Mediterranean Rivers (Gadiana River, SW Spain). Ecological Indicators 10: 1152-1161.
- Hesni, M.A.; Savari, A.; Sohrab, A.D. & Mortazavi, M.S. 2011. Gill histopathological changes in milkfish (*Chanos chanos*) exposed to acute toxicity of diesel oil. World Applied Sciences Journal 14: 1487-1492.
- Mallatt, J. 1985. Fish gill structural changes induced by toxicants and other irritants: a statistical review. Canadian Journal of Fisheries & Aquatic Sciences 42: 630-648.
- Mazon, A.F.; Cerqueira, C.C.C. & Fernandes, M.N. 2002. Gill Cellular Changes Induced by Copper Exposure in the South American Tropical Freshwater Fish *Prochilodus scrofa*. Environmental Research Section A 88: 52-63.
- Moitra, S.; Bhattacharjee, R. & Sen, N.S. 2012. Histopathological changes in the gills of air breathing teleost *Clarias batrachus* Linn. exposed to endosulfan. Asian Journal of Experimental Sciences 26: 23-26.
- Monteiro, S.M.; Rocha, E.; Fontainhas-Fernandes, A. & Sousa, M. 2008. Quantitative histopathology of *Oreochromis niloticus* gills after copper exposure. Journal of Fish Biology 73: 1376-1392.
- Mustafa, S.A.; Davies, S.J. & Jha, A.N. 2012. Determination of hypoxia and dietary copper mediated sub-lethal toxicity in carp, *Cyprinus carpio*, at different levels of biological organisation. Chemosphere 87: 413–422.
- Pandey, S.; Parvez, S.; Ansari, R.A.; Ali, M.; Kaur, M.; Hayat, F.; Ahmad, F. & Raisuddin, S. 2008. Effects of exposure to multiple trace metals on biochemical, histological and ultrastructural features of gills of a freshwater fish, *Channa punctata* Bloch. Chemico-Biological Interactions 174: 183–192.
- Patnaik, B.B.; Howrelia, H.J.; Mathews, T. & Selvanayagam, M. 2011. Histopathology of gill, liver, muscle and brain of *Cyprinus carpio communis* L. exposed to sublethal concentration of lead and cadmium. African Journal of Biotechnology 10: 12218-12223.
- Pelgrom, S.M.G.J.; Lock, R.A.C.; Balm, P.H.M. & Wendelaar Bonga, S.E. 1995. Integrated physiological response of tilapia, *Oreochromis mossambicus*, to sublethal copper exposure. Aquatic Toxicology 32: 303-320.
- Pereira, S.; Pinto, A.L.; Cortes, R.; Fontainhas-Fernandes, A.; Coimbra, A.M. & Monteiro, S.M. 2013. Gill histopathological and oxidative stress evaluation in native fish captured in Portuguese northwestern rivers. Ecotoxicology and Environmental Safety 90: 157-166.
- Santos, D.C.M.; Matta, S.L.P.; Oliveira, J.A. & Santos, J.A.D. 2011. Histological alterations in gills of *Astyanax* aff. *bimaculatus* caused by acute exposition to zinc. Experimental & Toxicologic Pathology 64: 861-866.
- Scardi, M.; Cataudella, S.; Di Dato, P.; Fresi, E. & Tancioni, L. 2008. An expert system based on fish assemblages for evaluating the ecological quality of streams and rivers. Ecological Informatics 3: 55-63.
- Stentiford, G.; Longshaw, M.; Lyons, B.; Jones, G.; Green, M. & Feist, S. 2003. Histopathological biomarkers in estuarine fish species for the assessment of biological effects of contaminants. Marine Environmental Research 55: 137-159.
- Teh, S.T.; Adams, S.M. & Hinton, D.E. 1997. Histopathologic biomarkers in feral freshwater fish populations exposed to different types of contaminate stress. Aquatic Toxicology 37: 51-70.
- Tlili, S.; Jebali, J.; Banni, M.; Haouas, Z.; Mlayah, A.; Helal, A.N. & Boussetta, H. 2010. Multimarker approach analysis in common carp *Cyprinus carpio* sampled from three freshwater sites. Environmental Monitoring and Assessment 168: 285–298.
- USEPA. 2007. Aquatic life ambient freshwater quality criteria – copper. 2007 Revision. EPA-822-R-07-

- 001 (CAS Registry Number 7440-50-8). Washington, DC: US Environmental Protection Agency and Office of Water.
- Van Dyk, J.C. & Pieterse, G.M. 2008. A histomorphological study of the testis of the sharptooth catfish (*Clarias gariepinus*) as reference for future toxicological assessments. *Journal of Applied Ichthyology* 24: 415-422.
- Viarengo, A.; Burlando, B.; Giordana, A.; Bolognesi, C. & Gabrielides, G.P. 2000. Networking and expert-system analysis: next frontier in biomonitoring. *Marine Environmental Research* 49: 483-486.
- Zhou, B. & Gitschier J. 1997. HCTR1: A human gene for copper uptake identified by complementation in yeast. *Proceedings of the National Academy of Sciences of the United States of America* 94: 7481-7486.
- Zorita, I.; Apraiz, I.; Ortiz-Zarragoitia, M.; Orbea, A.; Cancio, I.; Soto, M.; Marigo´mez, I. & Cajaraville, M. 2007. Assessment of biological effects of environmental pollution along the NW Mediterranean Sea using mussels as sentinel organisms. *Environmental Pollution* 148: 236-250.